

IN THE SPECIFICATION

(1) Delete the paragraph on page 19 lines 1-11 and replace it with:

A variety of options are available for measuring the amplification products as they are formed. One method utilizes labels, such as dyes, which only bind to double stranded DNA. In this type of approach, amplification product (which is double stranded) binds dye molecules in solution to form a complex. With the appropriate dyes, it is possible to distinguish between dye molecules free in solution and dye molecules bound to amplification product. For example, certain dyes fluoresce only when bound to amplification product. Examples of dyes which can be used in methods of this general type include, but are not limited to, SYBR® GREENER™ Syber GreenTM and PICOGREEN® Pico Green from Molecular Probes, Inc. of Eugene, Oreg., ethidium bromide, propidium iodide, chromomycin, acridine orange, Hoechst 33258, Toto-1, Yoyo-1, DAPI (4',6-diamidino-2-phenylindole hydrochloride).

(2) Delete the paragraph on page 73 line 25 to page 74 line 23 and replace it with:

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, the emulsifying agent CREMOPHOR EM™ Cremophor EM™ (BASF, Parsippany, N.J.) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, a pharmaceutically acceptable polyol like glycerol, propylene glycol, liquid polyethylene glycol, and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for

example, sugars, polyalcohols such as mannitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin. Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a polypeptide or antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

(3) Delete the paragraph on page 89 line to page 90 line and replace it with:

For relative quantitation of the distribution of PDE10A mRNA in cells and tissues the Perkin Elmer ABI PRISM® ABI Prism RTM 7700 Sequence Detection system or Biorad iCycler was used according to the manufacturer's specifications and protocols. PCR reactions were set up to quantitate PDE10A and the housekeeping genes HPRT (hypoxanthine phosphoribosyltransferase), GAPDH (glyceraldehyde-3-phosphate dehydrogenase), .beta.-actin, and others. Forward and reverse primers and probes for PDE10A were designed using the Perkin Elmer ABI PRIMER EXPRESS™ ABI Primer Express™ software and were synthesized by TibMolBiol (Berlin, Germany). The PDE10A forward primer sequence was: Primer1 (SEQ ID NO: 3). The PDE10A reverse primer sequence was Primer2 (SEQ ID NO: 5). Probe1 (SEQ ID NO: 4), labelled with FAM (carboxyfluorescein succinimidyl ester) as the reporter dye and TAMRA (carboxytetramethylrhodamine) as the quencher, is used as a probe for PDE10A. The following reagents were prepared in a total of 25 μ l: 1x TAQMAN® TaqMan buffer A, 5.5 mM MgCl₂, 200 nM of dATP, dCTP, dGTP, and dUTP, 0.025 U/ μ l of DNA polymerase AMPLITAQ GOLD™ AmpliTaq Gold™, 0.01 U/ μ l AmpErase AMPERASE® (uracil-N-glycosylase) and Probe1 (SEQ ID NO: 4), PDE10A forward and reverse primers each at 200 nM, 200 nM PDE10A FAM/TAMRA-labelled probe, and 5 μ l of template cDNA. Thermal cycling

parameters were 2 min at 50°C, followed by 10 min at 95°C, followed by 40 cycles of melting at 95°C for 15 sec and annealing/extending at 60°C for 1 min.

- (4) Insert after the claims the abstract on the page which follows.